

Production of mobile invertebrate communities on shallow reefs from temperate to tropical seas

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1 **Abstract**

2 Primary productivity of marine ecosystems is largely driven by broad gradients in
3 environmental and ecological properties. In contrast, secondary productivity tends to be more
4 variable, influenced by bottom-up (resource driven) and top-down (predatory) processes,
5 other environmental drivers, and mediation by the physical structure of habitats. Here, we use
6 a continental-scale dataset on small mobile invertebrates ('epifauna'), common on surfaces in
7 all marine ecosystems, to test influences of potential drivers of temperature-standardised
8 secondary production across a large biogeographic range. We found epifaunal production to
9 be remarkably consistent along a temperate to tropical Australian latitudinal gradient of
10 28.6°, spanning kelp forests to coral reefs (~3500 km). Using a model selection procedure,
11 epifaunal production was primarily related to biogenic habitat group, which explained up to
12 45% of total variability. Production was otherwise invariant to predictors capturing primary
13 productivity, the local biomass of fishes (proxy for predation pressure), and environmental,
14 geographic, and human impacts. Highly predictable levels of epifaunal productivity
15 associated with distinct habitat groups across continental scales should allow accurate
16 modelling of the contributions of these ubiquitous invertebrates to coastal food webs, thus
17 improving understanding of likely changes to food web structure with ocean warming and
18 other anthropogenic impacts on marine ecosystems.

19 **Keywords**

20 Macrofauna, epifauna, benthic ecosystems, trophic ecology, community ecology

21

22 **Introduction**

23 The production and transfer of biomass among constituents of an ecosystem is affected by a
24 diversity of processes that differ among scales. At local scales, biotic interactions such as
25 competition [1], predation [2] and facilitation or ecological complementarity (as related to
26 local species richness [3, 4]) influence productivity. In contrast, regional patterns in
27 productivity tend to relate to larger-scale variation in primary producer characteristics,
28 temperature, and nutrient availability (i.e. ‘bottom up’ processes; [5]). Reconciling these
29 varied drivers of community productivity has long been a goal of ecologists, particularly in
30 marine systems [6].

31 In this era of ‘big data,’ our capacity to simultaneously evaluate a suite of potential influences
32 has yielded novel insights regarding productivity – a fundamental ecosystem property [7].
33 Phytoplankton productivity, for example, can now be readily assessed across large
34 biogeographic scales using remote sensing tools [8-10]. However, secondary productivity—
35 particularly biomass production at the basal consumer level, including many small
36 heterotrophs that funnel energy through the food web—is less easily quantified, with
37 laborious field assessments generally required [11, 12]. For this reason, comparisons of
38 secondary productivity across broad biogeographic scales are relatively rare, and generalized
39 ecological and environmental drivers are yet to be identified (but see [13, 14]).

40 Reef ecosystems are among the most productive and diverse on earth. The productivity of
41 reefs is often quantified in terms of fish production [15], fisheries yield [16, 17], or the
42 primary productivity generated by phytoplankton or benthic algae [18]. A substantial
43 proportion of reef secondary production, though, is generated by small mobile invertebrates
44 (‘epifauna’) that inhabit the surfaces of macroalgae, coral, and other benthic structures [11,
45 19]. Epifauna are highly abundant, diverse and ubiquitous on shallow reefs worldwide, and
46 represent the main trophic link between benthic primary producers and small carnivores [20,

21]. Despite their fundamental role in coastal food webs, the drivers of epifaunal productivity—and thus, ‘fuel’ for most coastal food webs—have rarely been examined outside highly-controlled experiments [22, 23] and a few local- to regional-scale investigations [5, 13, 24].

Potential drivers of epifaunal productivity can be hypothesized based on documented patterns in other trophic groups and ecosystems, and on relationships described in previous studies of epifauna. Many biological processes are heavily influenced by temperature, and therefore strong latitudinal patterns in productivity are often reported. For example, in forests [25], open oceans [26], freshwater streams [14], and seagrass beds [27], productivity is generally highest at equatorial latitudes and lowest towards the poles, largely as a product of metabolic and growth rates scaling with temperature and light [28]. Concurrent spatial variation may also suggest unmeasured environmental factors, perhaps including evolutionary processes playing out over longer timeframes that favour more productive traits at low latitudes [29, 30]. Moreover, epifaunal secondary productivity may not respond as consistently as primary productivity to latitudinal temperature gradients. Although tropical/temperate differences have been observed [31], previous research indicates there may be no clear pattern in epifaunal productivity across smaller gradients or distinct locations [13, 32].

Both biotic (ecological) interactions and environmental drivers are fundamental determinants of food web structure and function [33], and their relative importance has been debated for several decades [6, 34]. Local-scale biotic interactions such as predation are clearly important in marine food webs [2, 22, 35, 36], and as such, variation in epifaunal productivity has often been discussed in terms of predation pressure [37-39]. Relationships between epifauna and various metrics of predation pressure, however, are inconsistent [22, 40]. Predation effects are further complicated by mesopredator release [41] and the fact that functional groups in addition to obligate invertivores, such as scraping and browsing herbivores, may ingest and

72 assimilate epifauna [42, 43], leading to greater trophic transfer along unexpected pathways.

73 The relationship between secondary productivity and biomass of potential predators may
74 therefore vary along large-scale gradients due to both the functional composition of predator
75 communities and the feeding behaviour within functional groups [44].

76 In concert with local-scale ecological interactions, broad-scale environmental drivers such as
77 changes in resource supply can equally influence secondary productivity. This phenomenon
78 may play out through changes in the abundance and composition of primary producers ,
79 which often correlate with changes in environmental conditions, for example light
80 (moderated by factors such as depth and turbidity in marine ecosystems; [13, 45]) and
81 nutrient availability [46]. Previous studies have indicated that food resources appear to set the
82 ceiling on total production of epifaunal communities after accounting for metabolic
83 contributions, with individuals redistributing along a size gradient to maximize community
84 productivity depending on whether they are exposed to predators [22].

85 Local-scale environmental drivers may also affect secondary productivity, albeit often via
86 interactions with local ecological processes or broad-scale environmental drivers. More
87 complex, stable and/or diverse habitats may support higher faunal productivity through
88 provision of greater abundance and diversity of food resources [11, 14, 47, 48], thus reducing
89 competition among secondary producers, or through increased protection from predation
90 [49]. Herbivorous amphipods often select more finely complex algal habitat based on the
91 quality of predation refugia, rather than the nutritional quality of the algae [50]. In addition,
92 while some algal species use chemical defences against fish herbivory, epifauna may be less
93 sensitive to these defences, selecting better-defended algal habitats as a refuge against
94 consumption by omnivores or herbivores [51]. Local-scale physical conditions – such as
95 wave energy and current flow in marine systems [52, 53] – and nutrients [54] or pollutants
96 [55], can all have substantial effects on faunal community structure and function. These

97 factors, and others such as removal of top predators [7, 56, 57], are often related to proximity
98 and density of human populations [58], and nearby industrial or agricultural activities [59,
99 60].

100 Here, we assembled a continental-scale dataset of shallow reef epifauna consistently surveyed
101 along the east coast of Australia, with the overarching aim of identifying major drivers of
102 variation in epifaunal secondary productivity across biogeographic provinces. Using multi-
103 model inference, we tested six hypotheses relating to expectations from ecological theory and
104 prior evidence (Table 1). We hypothesized that, like primary production, the major
105 constraints on local secondary production across large scales would be set by the amount of
106 resources and the abiotic environment, with smaller roles for biotic and other factors.

107 **Methods**

108 **Study area and field sampling**

109 Epifauna were sampled on shallow reefs at 11 eastern Australian locations, from southern
110 Tasmania (43.3°S) to Lizard Island in the northern Great Barrier Reef (14.7°S) (Fig. 1).
111 These locations represent a range of biogeographic regions, described in Appendix 1. A total
112 of 132 samples of diverse benthic microhabitats (comprising the most common biogenic
113 microhabitats available on rocky and coral reefs) and associated epifaunal invertebrates were
114 collected via SCUBA. Site selection, and sample collection and preservation follow protocols
115 described by Fraser, Stuart-Smith [61] and are presented in detail in Appendix 1.

116 **Laboratory processing and description of variables**

117 *Productivity estimates*

118 Preserved invertebrates from each sample were passed through a nested series of 12 sieves
119 stacked in descending order of mesh size, following a $\log_{\sqrt{2}}$ series (8, 5.6, 4.0, 2.8, 2.0, 1.4,
120 1.0, 0.71, 0.5, 0.355, 0.25, 0.18, 0.125 mm, after Edgar [62]). Invertebrates retained on each

121 sieve were washed into petri dishes and counted under a dissecting microscope, with data
122 binned by sieve mesh size.

123 Epifaunal abundance data by size bin were standardized to 1 m² planar seabed area (density)
124 prior to analysis following Fraser, Stuart-Smith [61]. Standardization by seabed area was
125 considered most appropriate for comparing epifaunal productivity to other trophic groups
126 such as fishes in food web models.

127 To calculate productivity, epifaunal biomass as ash-free dry weight (AFDW) of individuals
128 within each size bin was first derived from published estimates of mean biomass across
129 macrofaunal taxonomic groups [62]. Productivity estimates were calculated using the general
130 allometric equation given by Edgar [62]:

$$131 \quad P = (10^{(-2.31 + 0.8 * \log_{10}(B * 1000) + 0.89 * \log_{10}T)})/1000$$

132 where P is productivity of an individual (mg AFDW d⁻¹), B is the biomass of an individual
133 (mg AFDW), and T is water temperature (°C) at the time of sampling. Productivity estimates
134 of individual animals were then multiplied by density within each size bin, and size bin
135 productivity estimates summed to provide total productivity estimates (mg AFDW m⁻² d⁻¹)
136 for each sample. Productivity was calculated for a standardized temperature of 20°C
137 following Edgar [13], and hereafter referred to as P_{20} . The use of P_{20} is recommended to
138 eliminate the effects of temperature when investigating food webs, assuming that metabolic
139 and growth rates respond similarly to temperature change across trophic levels [13]. We note
140 that this method for estimating biomass and productivity was originally established for
141 individuals ≥ 0.5 mm; here we assume the equations used by Edgar [62] also apply to smaller
142 individuals (≥ 0.125 mm) based on linear extrapolation of well-supported trends (i.e. R^2
143 ranging from 0.87 to 0.98 [67]). In Appendix 1 we elaborate on methods used for
144 productivity estimates for samples collected using the venturi air-lift (i.e. from massive corals

145 and turfing algae) and on methods used to visualize variation in epifaunal productivity among
146 sampling locations.

147 *Predictor variables*

148 Predictor variables and the models in which they are applied are summarized in Table 1,
149 while details of predictor variables are provided in Appendix 2. Appendix 1 presents detail on
150 how and from where data were collected for each predictor variable.

151 **Data analyses**

152 Estimated epifaunal P_{20} per m^2 of seabed (estimated by multiplying the fraction of benthic
153 cover provided by each microhabitat within each site by the estimated P_{20} associated with that
154 microhabitat) was averaged within each of the 11 sampling locations to give mean P_{20} (mg
155 AFDW $m^{-2} d^{-1}$) for each location. These data were plotted against latitude using a linear
156 model in R (R Development Team 2017).

157 Six hypotheses were tested using multiple regression models parameterized with the
158 appropriate predictors (Table 1) in a multi-model inference framework [63] (see [64] for the
159 dataset and R code used for analysis). We fit a separate linear model to \log_{10} transformed P_{20}
160 (per m^2 of individual microhabitat sampled) to test each hypothesis with the set of associated
161 predictor variables using the full (not summarized per location) dataset ($n = 115$) (Table 1).
162 Assumptions of each model were tested using variance inflation factors (VIF) for
163 independence of predictors and residuals were examined to ensure normality. We then used
164 Akaike information criterion with small sample correction (AICc) to evaluate the likelihood
165 of each model. We selected the best-supported model based on the Akaike weight, which
166 describes the relative likelihood of each model given the candidate set of models. The Akaike
167 weight (AICwt) ranges from 0-1, with 0 being no support and 1 being total support [63]. The
168 best supported models were further evaluated by Type-III ANOVA using the *car* package

169 [65] and Tukey post-hoc comparison of means. We fit the models using R version 3.6.3 [66]
170 and used the *AICcmodavg* package to compute Akaike weights [67].

171 Analyses described above were also conducted using temperature-dependent productivity
172 (results presented in Table S1). However, since modelling temperature-dependent
173 productivity as a function of temperature could lead to mathematical dependence between the
174 response and the predictor, P_{20} was chosen as the preferred response variable.

175 **Results**

176 Across 28.6 degrees of latitude, we found little variation in total epifaunal community
177 productivity (P_{20} ; mg AFDW $m^{-2} d^{-1}$), at both the individual sample level and the location
178 level based on the contribution of different microhabitats to total benthic cover (Fig. 2a). The
179 lack of variation in productivity standardized by temperature (P_{20}) with latitude indicates that
180 epifaunal productivity should maintain similar productivity relativities with other food web
181 elements (e.g. fishes, primary producers), all equally varying with temperature as predicted
182 by metabolic theory.

183 The habitat group model was overwhelmingly the best supported model to explain variation
184 in epifaunal P_{20} (AICwt = 0.96; Table 1), suggesting that epifaunal secondary productivity is
185 predominantly driven by characteristics of the immediate habitat group occupied by an
186 assemblage (i.e. macroalgae, live coral, sessile invertebrate, or turfing algae). The
187 microhabitat model, which includes finer but more numerous microhabitat categories than the
188 habitat group model, was supported to a much lesser degree (AICwt = 0.04), suggesting that
189 the explanatory power gained by this increased resolution was not worth the loss of additional
190 degrees of freedom, while all other hypotheses had no support according to their Akaike
191 weights (Table 1).

192 Within the habitat group model, epifaunal P_{20} differed significantly among habitat groups (F -
193 value = 19.4, $P < 0.001$; Fig. 2b; Table S2). Tukey pair-wise comparison of mean P_{20} among
194 habitat groups indicated significant differences between macroalgae and live coral ($P =$
195 0.0033), and between turfing algae and live coral ($P = 0.010$). Epifaunal P_{20} also showed a
196 significant positive correlation with branching (F -value = 6.3, $P = 0.011$; Fig. 3a; Table S2).
197 However, the effect of branching varied significantly among habitat groups (F -value = 3.3, P
198 = 0.024; Table S2), with the overall positive correlation between branching and P_{20} largely
199 driven by macroalgae and turfing algae habitat groups (Fig. 3a).

200 Our model selection analysis suggests that the near constant epifaunal productivity observed
201 on reefs along the east coast of Australia is a product of trade-offs in the dominant habitat
202 groups across the latitudinal gradient (Fig. 4). Moving from tropical to temperate latitudes,
203 the loss of live coral and associated secondary productivity is compensated by increased
204 contributions by communities of epifauna inhabiting turfing algae and sessile invertebrate
205 habitat groups, while macroalgal communities remain reasonably constant across the entire
206 latitudinal range.

207 **Discussion**

208 Ecosystem productivity has historically been considered to be predominantly a function of
209 environmental drivers that regulate the availability of resources [6, 7, 68]. Here, we find that
210 habitat group primarily determines the degree of secondary productivity provided by small
211 marine invertebrates to shallow reef food webs. Trade-offs in the local productivity afforded
212 by each of four broad habitat groups (corals, macro- and turfing algae, and sessile
213 invertebrates) led to a remarkably consistent trend in epifaunal secondary productivity from
214 temperate to tropical zones.

215 While community structure and function have long been viewed through the lens of resource
216 control, the controlling resource has often been framed in terms of biomass and energy
217 transfer among trophic groups (i.e. carbon acquisition) [7, 13, 69, 70]. However, niche theory
218 also acknowledges space as an important resource (i.e. the ‘Hutchinsonian’ niche), harkening
219 back to seminal contributions on the organization of sessile organisms in rocky intertidal
220 ecology [71, 72]. Habitat resources, additional to food resources, appear responsible for
221 large-scale patterns in epifaunal community structure [61, 73]. This seems also to be the case
222 in the current study with regards to their production, echoing a recent finding in communities
223 of freshwater stream invertebrates in North America [14].

224 **Why is habitat so important?**

225 Several potential mechanisms may explain our finding. First, while epifaunal assemblages
226 comprise a diversity of functional groups, herbivores (the ‘mesograzers’) typically dominate
227 [13, 74]. Mesograzers tend to rely on microphytobenthic films and filaments, with some
228 larger animals consuming macroalgae [75]. Macroalgal habitats present abundant food
229 resources in the form of microphytobenthos and host algal tissue, potentially facilitating
230 greater productivity of epifauna than habitats without these resources [35]. Filamentous
231 turfing algae, in addition to offering a direct food source for mesograzers, tends to host
232 microalgal films and capture high volumes of detritus [76], presenting an abundance and
233 diversity of trophic resources for different epifaunal functional groups [75]. By contrast, live
234 hard coral offers minimal food for herbivorous mesograzers [77, 78], making it largely food
235 resource-poor except for particles trapped by coral polyps and the coral mucus consumed by
236 some larger decapod taxa [79]. Epifauna selecting soft coral and sponge habitats, comprising
237 the sessile invertebrate habitat group, are likely to encounter fewer food resources. Soft corals
238 use allelopathic defences to resist colonisation by microphytobenthos and epiphytes, and

239 consumption by epifauna [80]. Sponge tissue is consumed by some epifauna, however most
240 sponge-dwellers consume external food sources [81, 82].

241 Variation in epifaunal productivity may also be influenced by differential predation
242 susceptibility among benthic habitats. Habitat structural complexity and its role in shaping
243 predator-prey relationships has long been discussed [49, 78, 83, 84], and may be a factor
244 determining the relationship between epifaunal productivity and habitat groups. Epifaunal
245 productivity increased with our metric of habitat complexity (degree of branching) (Fig. 3a),
246 presumably due to the added protection from predators offered by more complex habitat [84,
247 85]. However, the degree to which this benefit is realized depends greatly on the habitat type
248 (Fig. 3a, 3b). For example, macroalgal habitat was the most highly branched and supported
249 among the highest estimates of epifaunal productivity, however live coral was also highly
250 branched but supported the least productive epifaunal assemblages.

251 This apparent inconsistency raises the question of whether physical complexity provides
252 actual or perceived refuges for epifaunal prey [78] and may be partly resolved by considering
253 the scale at which complexity is quantified. While live branching coral is complex at scales
254 ranging from millimetres to centimetres, the complexity of turfing algae is at a sub-millimetre
255 to millimetre scale, and macroalgae complexity ranges from sub-millimetre through to
256 centimetres [31]. In studies comparing macroalgae species [86] or artificial algal habitats of
257 differing complexity [87], small invertebrates generally select more finely complex habitat
258 that offers predation refugia appropriate for the invertebrate body sizes. Macroalgae
259 complexity can also be finely partitioned by much larger herbivorous fishes [88]. If
260 microhabitat complexity were quantified to higher resolution, for example by using fractal
261 dimensions [89], stronger relationships between epifaunal productivity and habitat
262 complexity would perhaps be evident, as would consistency between the complexity of
263 habitat groups and the productivity they support.

264 In addition to complexity, predation pressure may vary as a result of particular characteristics
265 of the different habitat groups. For example, while the heterotrophy of hard corals largely
266 involves the consumption of zooplankton [90, 91], small epifaunal invertebrates could fall
267 prey to coral polyps. Hard corals also often use physical defence strategies, such as ‘sweeper
268 tentacles’, to resist colonization by small epiphytes and epifauna [92]. In addition, the rigid
269 structure of branching hard coral limits the ability of mobile invertivores (e.g. fish) to
270 penetrate the habitat in order to extract epifaunal prey [93]. Hence, branching coral can
271 provide refugia for larger epifaunal invertebrates that may be less susceptible to consumption
272 by coral polyps [39, 73, 94].

273 Fish communities on tropical reefs have been shown to comprise proportionally more
274 herbivores compared with temperate reefs, which support more omnivorous fishes, while
275 invertivores are consistently common across all latitudes [95]. While total fish biomass is
276 used here as a proxy for predation pressure, understanding the differences in predation
277 exposure for epifauna among different microhabitats would require more detailed study of the
278 functional composition and feeding behaviour of local fish communities. For example,
279 predation by omnivores or consumption of epifauna by herbivores may vary among algal
280 microhabitats depending on chemical defences against fish herbivory or the palatability of
281 algae, as epifaunal invertebrates may be insensitive to chemical defences [51] or choose less
282 palatable algal microhabitats based on refuge quality [50].

283 Interestingly, neither site-scale estimates of predator biomass, nor temperature or primary
284 productivity (assessed using water column chlorophyll content as a proxy) appeared to be
285 explicitly related to variation in epifaunal productivity. Our use of P₂₀ controls for a major
286 environmental factor, temperature, although theory and recent studies suggest that
287 temperature effects are most likely to manifest through enhancing the (consumable) resource
288 base, rather than acting directly on community production [14, 96, 97]. Metabolic rate scales

289 with temperature at approximately similar rates across trophic levels, resulting in
290 proportionally similar production/temperature changes [13]. Given that habitat group affects
291 potential food resources available for epifauna, whereas temperature had little apparent
292 influence on secondary productivity, our results are consistent with the hypothesis that
293 epifaunal productivity is limited predominantly by food resource ceilings [13, 22].

294 **Ecological implications**

295 Epifaunal invertebrates are extremely prolific in coastal and shallow reef ecosystems, with a
296 very high proportion of their biomass consumed by larger invertebrate predators and fishes
297 [11]. Consequently, epifaunal communities comprise a critical basal component in shallow
298 marine food webs [85]. Understanding the factors that promote productive epifaunal
299 communities is crucial for the goal of ensuring high trophic transfer and food web stability
300 for coastal and shallow reef ecosystems. Given that the biotic habitat group occupied by the
301 epifaunal assemblage was here found to explain >45% of the variance in secondary
302 productivity along an extensive biogeographic gradient, understanding changes to benthic
303 habitat group availability is the critical first step to achieving this goal.

304 In selecting microhabitats to sample, we attempted to include all common types of biogenic
305 cover found on shallow coral and rocky reefs in eastern Australia. However, direct
306 anthropogenic stressors, combined with climate change, are shifting the distribution and
307 abundance of biogenic habitat groups common to rocky and coral reefs [98-100]. Our results
308 reveal an important indirect pathway for the effects of global, regional, and local scale
309 environmental changes to alter reef ecosystems. Ocean temperature has been identified as the
310 most important driver of the benthic composition of biogenic habitat groups on both rocky
311 and coral reefs [101]. Other important drivers include human population density, nutrient
312 availability, wave exposure, and the density of habitat-transforming fauna such as

313 herbivorous sea urchins or corallivorous crown-of-thorns sea stars [101-103]. Turf and
314 sometimes macroalgae are succeeding corals lost to bleaching and other local stressors [98,
315 104, 105]. Macroalgae beds on rocky reefs are declining in many regions [106], often to be
316 replaced by turf as oceans warm and voracious herbivores undergo range extensions and
317 population outbreaks [100, 107, 108].

318 Mediated by shifts in available reef habitat groups, these drivers can potentially affect
319 epifaunal invertebrate communities and food web processes. Our results imply changes to
320 epifaunal secondary productivity should be predictable if habitat group transformation is well
321 documented or accurately predicted. Replacement of live coral by turfing algae or
322 macroalgae will likely increase epifaunal secondary productivity on tropical and subtropical
323 reefs (Fig. 4) [94]. If turf replaces macroalgae on temperate reefs, a significant increase in
324 epifaunal productivity may be expected, whereas the succession of subtropical macroalgae by
325 turf is likely to result in minimal change (Fig. 4). Rather, relatively high epifaunal
326 productivity may be maintained on subtropical reefs, as turfing and macroalgae both support
327 similarly highly productive assemblages of epifaunal invertebrates.

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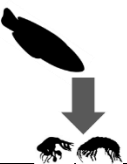


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632 **Tables**

633 **Table 1** The hypotheses (epifaunal community P_{20} is predominantly driven by: H1 – H6) and
 634 linear models tested to explain variation in epifaunal P_{20} , with predictions (P) included within
 635 models. Partial R^2 indicates the proportion of variance explained by each predictor within
 636 models; multiple R^2 indicates the raw unadjusted R^2 for each model. Model selection was
 637 based on the Akaike weight, which describes the relative likelihood of each model given the
 638 set of candidate models.

Hypothesis (H)	Model and Predictions (P)	Partial R^2	Multiple R^2	Akaike weight
H1 – Predation pressure 	PREDATION MODEL		0.032	<0.01
	P1 – P_{20} declines with increased total fish biomass	0.004		
	P2 – P_{20} declines with increased cryptic fish abundance	0.029		
H2 – Resource availability 	RESOURCE MODEL		0.122	<0.01
	P3 – P_{20} declines as depth increases (reducing light)	0.031		
	P4 – P_{20} increases with epiphyte load	0.048		
	P5 – P_{20} increases with chlorophyll-a	0.005		
H3a – Characteristics of immediate habitat, fine microhabitat scale 	MICROHABITAT MODEL		0.594	0.04
	P7 – P_{20} varies significantly among microhabitats	0.548		
	P8 – P_{20} increases with habitat branching/complexity	0.025		
H3b – Characteristics of immediate habitat, coarse habitat group scale	HABITAT GROUP MODEL		0.450	0.96
	P10 – P_{20} varies significantly among habitat groups	0.344		
	P11 – P_{20} increases with habitat branching/complexity	0.030		
	P12 – the effect of branching on P_{20} varies among habitat groups	0.069		

	P13 – P ₂₀ increases with the maximum length of habitat	0.007		
H4 – Local environmental factors 	ENVIRONMENT MODEL		0.114	<0.01
	P6 – P ₂₀ increases with mean SST	0.049		
	P14 – P ₂₀ declines with increased wave exposure	0.036		
	P15 – P ₂₀ declines with increased relief	0.009		
	P16 – P ₂₀ declines with increased slope	0.014		
	P17 – P ₂₀ declines with increased current strength	0.006		
H5 – Geographic location 	SPATIAL MODEL		0.091	<0.01
	P18 – P ₂₀ declines towards higher latitudes	0.054		
	P19 – P ₂₀ varies significantly with longitude	0.037		
H6 – Human population impacts 	HUMAN IMPACTS MODEL		0.077	<0.01
	P20 – P ₂₀ increases with human population density	0.077		

640 **Figure legends**

641 **Fig. 1** Map of eastern Australia showing sampling locations, sampling dates and number of
642 sites.

643

644 **Fig. 2** Linear regression (**a**) of mean \log_{10} total epifaunal community daily productivity (P_{20})
645 against latitude. The large black points represent mean P_{20} within each of the 11 sampling
646 locations, estimated by multiplying the fraction of benthic cover provided by each
647 microhabitat within each site by the estimated P_{20} associated with that microhabitat; the black
648 line represents the regression of those data against latitude. The small grey points represent
649 epifaunal P_{20} for individual samples; the grey line represents the regression of those data
650 against latitude. Grey shading represents 95% confidence intervals.

651 Box plots (**b**) of variation in \log_{10} epifaunal assemblage P_{20} among habitat groups. Horizontal
652 lines in each box plot represent third quartile, median and first quartile. The whiskers extend
653 to 1.5 x interquartile range. Dots represent outliers. Asterisks indicate significant differences
654 between habitat group pairs (* P <0.05; ** P <0.01).

655

656 **Fig. 3** Linear regression (**a**) of mean \log_{10} epifaunal P_{20} against microhabitat degree of
657 branching, with colors indicating habitat groups, and black line the overall mean. Higher
658 branching equates to higher complexity and translates to higher productivity on average.
659 Points represent individual samples; grey shading represents 95% confidence interval of
660 overall mean. Horizontal boxplots (**b**) show variation in the degree of branching within each
661 habitat group. Vertical lines in each box plot represent third quartile, median and first
662 quartile. The whiskers extend to 1.5 x interquartile range.

663

664 **Fig. 4** Mean \log_{10} epifaunal P_{20} associated with each habitat group across four climatic zones
665 within the latitudinal gradient sampled. Mean P_{20} among habitat groups is represented for
666 each climatic zone by the bar titled 'All'. Climatic zones represent the following latitudinal
667 ranges: cool temperate (-43.3 to -37.7°S), warm temperate (-37.6 to -31.9°S), subtropical (-
668 31.8 to -26.1°S), tropical (-20.4 to -14.6°S).

669