

1 Running head: Shelf stability of Neutral Electrolysed Water

2 Research Paper

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4 Effect of storage conditions on shelf stability of undiluted neutral electrolysed water

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24 **Abstract**

25 Neutral electrolysed water (NEW) is an oxidising sanitiser that can be made locally on-site but is  
26 often stored in ready-to-use format in order to accumulate the large volumes required for periodic  
27 or seasonal use. The shelf stability of NEW sanitiser was therefore assessed under various storage  
28 conditions to guide the development of protocols for its industrial application. To that end, fresh  
29 NEW with an available chlorine concentration (ACC) of 480 mg/L, pH 6.96 and oxidation reduction  
30 potential (ORP) of 916 mV was stored under different conditions. These were open or sealed  
31 polypropylene bottles, three different surface area-to-volume (SA:V) ratios (0.9, 1.7 and 8.7) and two  
32 temperatures (4, and 25 °C). NEW stored at 4 °C was significantly more stable than NEW stored at  
33 25 °C, where ACC and pH decreased by 137 mg/L and 0.7, respectively, while ORP increased by 23  
34 mV after 101 days of storage. At 25 °C, ACC decreased to <0.01 mg/L after 52 days in bottles with a  
35 SA:V ratio of 8.7 with a similar decrease after 101 days in bottles with a SA:V ratio of 1.7. However,  
36 pH decreased by up to 3.7 pH units but ORP increased by up to 208 mV. Antimicrobial efficacy of  
37 'aged' electrolysed oxidising (EO) water with different ACC and ORP, but the same pH (i.e.,  $3.4 \pm 0.2$ )  
38 was evaluated against *Escherichia coli* and *Listeria innocua* to determine any differences in residual  
39 antimicrobial activity. EO water with an ACC of  $\geq 7$  mg/L and ORP of 1,094 mV caused at least a 4.7-  
40 log reduction, whereas EO water with non-detectable ACC and considerably high ORP (716 mV) had  
41 little antimicrobial effect (<1-log reduction). Results from this study indicate that the efficacy of  
42 NEW as a sanitiser for large scale applications like horticulture can be maintained for at least 3  
43 months when stored in closed containers with low SA:V ratio at low temperatures.

44

45 **HIGHLIGHTS**

- 46 • NEW sanitiser (480 mg/mL) was more stable to storage at 4 °C than at 25 °C.
- 47 • Containers with a lower SA:V ratio improved the stability of NEW.
- 48 • Antimicrobial effects of NEW were significant at chlorine concentration > 7 ppm.

49

50 Electrolysed water, referred to as electrolysed oxidising (EO) water, is a non-specific broad-  
51 spectrum sanitiser (28). It is typically generated by the electrolysis of pure water with added  
52 hydrochloric acid (HCl) and/or NaCl in an electrolytic cell (3). Water molecules and chloride ions can  
53 then be electrolytically reacted to form different chlorine species such as hypochlorous acid (HOCl),  
54 hypochlorite ions ( $\text{ClO}^-$ ) and chlorine ( $\text{Cl}_2$ ). The proportion of these species in EO water is mainly  
55 dependent upon the pH of the EO water generated (28). Accordingly, different types of EO water  
56 exist and can be produced by changing the composition of the feed stream, the reaction conditions,  
57 and the arrangements of the electrolytic cell. These include acidic, slightly acidic, neutral and  
58 alkaline EO water. Among all types, neutral EO water (NEW) has gained increasing attention. The  
59 neutral pH minimises corrosion potential and is physiologically compatible. The pH of NEW also  
60 minimises safety issues from  $\text{Cl}_2$  off-gassing and maximises the availability of the hypochlorous acid  
61 species and therefore maximises antimicrobial efficacy (22).

62 EO water has notable biocidal activity, which is mainly due to its physicochemical properties such  
63 as available chlorine concentration (ACC), pH and oxidation reduction potential (ORP) (2, 22). There  
64 are numerous studies demonstrating that EO water could be used as a surface disinfectant of  
65 different materials commonly found in food processing facilities including cutting boards (17),  
66 stainless steel, vitreous china, ceramic tiles, and glass (21). Other studies have also reported their  
67 potential as an effective sanitiser for a range of fresh fruits and vegetables (12-14), barley grains  
68 (10), and a variety of meat and produce (5, 9, 21). Reviews by Huang *et al.*, (10) and Rahman *et al.*,  
69 (22), indicated that the antimicrobial efficacy of EO water can range from no effects to at least 6-log  
70 reduction in viable count. This variation has been suggested to be related to both intrinsic and  
71 extrinsic factors, including the type of EO water used, temperature, ACC, food composition and  
72 surface characteristics, water hardness, organic load, and bacterial type (10). Therefore,  
73 understanding these factors, their interactions, and how they affect the antimicrobial effects of NEW  
74 would aid its development as an effective and reliable antimicrobial application in the food industry.

75 In addition to the need to better characterise the factors affecting EO efficacy, improved studies  
76 to determine the best storage conditions for EO water are important. In horticulture, produce that is  
77 eaten raw requires sanitation after harvest and possibly in the field pre-harvest to moderate the  
78 presence of pathogens in harvested produce and/or control phytopathogens that can reduce crop  
79 yield. These large-scale applications require large volumes of sanitiser, which may be mixed or made  
80 on-site or made in ready-to-use format and transported to the point of application. EO water is  
81 typically produced in an electrolytic cell and accumulated in a holding vessel until used. EO water  
82 once produced, needs to be stored under the conditions that best preserve its stability as a sanitizer.

83 Previous studies have identified a range of storage factors including material type, light  
84 conditions, temperature, agitation level, and opened or closed container to influence the shelf  
85 stability of EO water (3, 11, 23, 24, 32, 33). However, these studies examined EO water with a low  
86 ACC (i.e.,  $\leq 100$  mg/L). Such low levels of ACC do not reflect the large-scale application of NEW as a  
87 sanitiser in commercial settings, in which ACC  $> 400$  mg/L is commonly produced (7, 18, 15).  
88 Arguably, freshly made NEW with a higher initial ACC may result in longer shelf stability but this  
89 remains unclear.

90 The aim of this study was, therefore, to assess the changes in physicochemical properties (i.e.,  
91 ACC, pH, and ORP), of NEW at a high level of ACC (i.e., that is relevant to commercial use, initial ACC  
92 of 480 mg/L, pH 6.96, ORP 916 mV) stored under various conditions. Another aspect of this study  
93 was to specifically characterise the antimicrobial efficacy of 'aged' EO water (with different ACC and  
94 ORPs at the same pH) to provide an insight into its potential mode of action and an assurance that  
95 degradation mechanisms or products do not alter the relative efficacy of the antimicrobial activity.

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## MATERIAL AND METHODS

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100

**Preparation of electrolysed water solution.** NEW was generated by the electrolysis of a saturated sodium chloride solution using an Envirolyte<sup>®</sup> ELA-400 (Envirolyte Industries International OÜ, Tallinn, Estonia) (4, 22). ACC (i.e. free chlorine) was analysed colorimetrically, using a Compact

101 CLO2+ meter (Palintest, Australia; with a detection limit of 0.01 mg/L), which determines ACC by a  
102 colourimetric reaction of free chlorine with di-ethyl-p-phenyl diamine (DPD). pH and ORP were  
103 analysed with an Orion 250A pH meter (Orion, USA) and a MW 500 ORP meter (Milwaukee, USA),  
104 respectively. The analyte produced had an initial ACC, pH and ORP of 480 mg/L, 6.96 and 916 mV,  
105 respectively.

106 **Effect of storage conditions on stability of NEW.** The changes in physicochemical properties of  
107 NEW were measured when stored under two different conditions. These included: (i) small volume  
108 bottles (50, 250 and 500 mL) with container surface area-to-volume ratios (SA:V ratios) of 8.7, 1.7,  
109 and 0.9 stored at 4 and 25 °C; and (ii) opened or closed bottles (500 mL) stored at 25 °C (with a SA:V  
110 ratio of 0.9). The surface area-to-area volume ratio was determined by the surface area of a  
111 cylindrical bottle (436.4 cm<sup>2</sup>) divided by the volume of each respective solution (Fig. S1). All  
112 experiments were undertaken in the dark. This is because light is well known to decompose HClO  
113 and hypochlorite anions, which absorb energy in the region of 292 – 380 nm [24-26].

114 For Experiment One, NEW was produced and dispensed into 25 L polyethylene containers  
115 and stored sealed at 4 °C for 1 day. NEW was transferred into 18 x 500 mL, wide-mouth  
116 polypropylene screw cap bottles (LP1403PP-500, Wiltronics, VIC, Australia), which were tightly  
117 capped and stored at 4 or 25 °C in the dark (three replicates per treatment of 50, 250 and 500 mL  
118 with SA:V ratios of 8.7, 1.7, and 0.9 respectively, stored at either 4 or 25 °C). For each treatment, the  
119 ACC, pH and ORP of the NEW was measured periodically from the same bottles over a period of  
120 storage up to 101 days.

121 For Experiment Two, NEW was produced and dispensed into 25 L polyethylene containers  
122 and transferred into 42 x 500 mL polypropylene screw cap bottles (three replicates per treatment  
123 per time point). In the first treatment 'closed' bottles were filled with NEW such that there was no  
124 head space and were tightly capped. For the second treatment: 'opened' bottles were filled, and the  
125 mouth of the bottles were covered loosely with aluminium foil. All bottles were stored in the dark at

126 25 °C. For each treatment, ACC, pH and ORP were measured periodically from different bottles over  
127 a period of storage up to 62 days.

128

129 **Evaluating antimicrobial efficacy of aged EO water.** *Escherichia coli* K12, MG1655 and *Listeria*  
130 *innocua*, ATCC33090 were used to assess the antimicrobial efficacy of the EO water after storage.  
131 Cultures were obtained from the culture collection of the Tasmanian Institute of Agriculture,  
132 University of Tasmania, Australia.

133

134 **Preparation of bacterial inocula.** Bacterial cultures, previously maintained at -80 °C, were  
135 resuscitated by streaking onto Brain-Heart Infusion (BHI) (AM11, AMYL, VIC, Australia) agar and  
136 incubated at 37 °C for 24 hours. A well-isolated single colony from each culture was aseptically  
137 transferred into 10 mL of BHI broth and incubated at 37 °C for 24 hours to achieve a cell density of  
138 approximately 10<sup>9</sup> CFU/mL. The cultures were stored at 4°C and used as 'stock' cultures within a  
139 week.

140 To prepare the inocula, stock cultures of *E. coli* and *L. innocua* were diluted 1:10<sup>3</sup> in 10 mL of  
141 BHI broth to prepare working cultures. These cultures were incubated at 37 °C for 20 hours to  
142 achieve a cell density of approximately 10<sup>9</sup> CFU/mL. A 1 mL aliquot of each of these working cultures  
143 was then harvested by centrifugation at 5,000 *g* for 5 minutes (Eppendorf 5417R) at 20 °C. The pellet  
144 was resuspended and washed twice with 1 mL of 0.1% peptone water (LP0037, Oxoid LTD.,  
145 Basingstoke, Hampshire, England) with subsequent centrifugation at 5,000 *g* for 5 minutes at 20 °C.  
146 The final washed pellet was resuspended in 1 mL of 0.1% peptone water and used as the inoculum  
147 (approximately 10<sup>9</sup> CFU/mL) for trials to assess antimicrobial efficacy.

148

149 **Antimicrobial evaluation.** EO water with different SA:V ratios that had been stored at 25 °C for  
150 87 and 101 days in Experiment One was used to specifically study the antimicrobial effects of 'aged'  
151 EO water. After measurement of ACC, ORP and pH, an aliquot (0.9 mL) of the aged EO water was

152 transferred into a sterile microcentrifuge tube and combined with 0.1 mL of either an *E. coli* or *L.*  
153 *innocua* suspension outlined above. Bacterial numbers, before and after exposure to treatments for  
154 1, 5 and 10 minutes at approximately 20 °C, were determined by spread plating 0.1 mL aliquots of  
155 appropriately diluted samples on BHI agar. After incubation at 37 °C for 18 hours, bacterial colonies  
156 were enumerated.

157 **Statistical analysis.** All experiments were set up in a completely randomised design with three  
158 replications per treatment. The effects of temperature, opened/closed storage, volume, time and  
159 their interactions were analysed by two-way analysis of variance (ANOVA) within a repeated  
160 measures framework using a Kenward-Rogers degrees of freedom adjustment. For the antimicrobial  
161 study, treatment effects were analysed by one-way ANOVA for each exposure time. The  
162 assumptions of ANOVA, such as homogeneity of variance and the Gaussian distribution, were  
163 confirmed by the use of quantile-quantile plots and residual plots for all variables. Significant  
164 differences were established at a level of  $P < 0.05$ . All statistical analyses were done using SAS version  
165 9.3.

## 166 RESULTS

167 **Effect of surface area to volume ratio and temperature on the stability of NEW.** pH and ORP,  
168 but not ACC, were significantly influenced by the interaction between volume, temperature and time  
169 (Table S1). When NEW was stored at 4°C for 101 days, the ACC decreased for all three SA:V ratios  
170 (Fig. 1a). The observed decrease also occurred faster with an increase in SA:V. The three treatments  
171 (SA:V ratios of 8.7, 1.7, and 0.9) had a final ACC of 343, 400, and 420 mg/L, respectively (Fig. 1a). The  
172 initial pH of 7.0 remained relatively stable, decreasing to 6.5, 6.4, and 6.3, for treatments with SA:V  
173 ratio of 8.7, 1.7, and 0.9, respectively (Fig. 1b). The ORP also remained relatively stable throughout  
174 the duration of the experiment. From an initial ORP of 916 mV, final ORP levels were 955, 961, and  
175 939 mV for SA:V ratios of 8.7, 1.7, and 0.9, respectively (Fig. 1c).

176 When stored at 25°C, the ACC decreased from 480 mg/L to <0.01 mg/L for bottles with a SA:V  
177 ratio of 8.7 and 1.7 by day 53 and 101, respectively (Fig. 1a). This was in contrast to bottles with a  
178 SA:V ratio of 0.9 in which the ACC decreased from 480 to 59 mg/L over 101 days (Fig. 1a), reinforcing  
179 the influence of the SA:V ratio on retention of ACC. The pH of all NEW treatments declined over the  
180 first 65 days of storage and then remained relatively stable. Bottles with SA:V ratios of 8.7, 1.7, and  
181 0.9 had final pH of 3.6, 3.2, and 3.3, respectively (Fig. 1b). The ORP of all treatments initially  
182 increased from 916 to 1,130 mV within the first 52 days (Fig. 1c). This was followed by a period in  
183 which the ORP of all bottles, except for the bottles with a SA:V ratio of 8.7, remained constant above  
184 1,100 mV. On days 31 and 80, the ORP of NEW in bottles with a SA:V ratio of 8.7 and 1.7 decreased  
185 from above 1,000 mV to 493 mV and 716 mV, respectively.

186

187 **Effect of opened or closed containers on the stability of NEW at 25 °C.** In these experiments,  
188 ACC, pH and ORP were significantly influenced by the interaction between treatment and time  
189 (Table S2). The ACC declined more quickly in opened bottles when compared to closed bottles (Fig.  
190 2a). The initial ACC of 480 mg/L for both opened and closed treatments decreased to 170 and 253  
191 mg/L over the course of 62-day storage, respectively. The pH of NEW with both opened and closed  
192 treatments decreased from 7.0 to 4.8 and 3.6, respectively (Fig. 2b). For the ORP data, there was an  
193 increase in ORP for both treatments within the first 50 days of storage (from 916 mV to  
194 approximately 1,100 mV; Fig. 2c). This was followed by a period in which their ORP appeared to be  
195 unchanged.

196

197 **Antimicrobial efficacy of aged EO water.** Aged EO water with different ACC and ORP, but at the  
198 same pH (pH 3.4±0.2), was found to significantly influence the mean log reduction of *E. coli* and *L.*  
199 *innocua* for each exposure time (Table 1; P <0.05). The initial inoculum level of *E. coli* and *L. innocua*  
200 was 9.09 and 9.21 log CFU/mL, respectively. EO water with an ACC of 63 mg/L, and ORP of 1,121 mV  
201 reduced *E. coli* and *L. innocua* by to below the detection limit following one minute of exposure. EO



202 water with an ACC of 7 mg/L and ORP of 1,094 mV, however, reduced *E. coli* only by  $4.76 \pm 0.09$  log  
203 CFU/mL within one minute. Thereafter, no further effects were observed with longer exposure  
204 times. For EO water with non-detectable ACC (<0.01 mg/L) and ORP of 716 mV, reduced *E. coli* and  
205 *L. innocua* by  $0.98 \pm 0.08$  log CFU/mL and  $1.02 \pm 0.02$  log CFU/mL, respectively after one minute with  
206 no further effects observed after 10 minutes. This was similar to our observation for EO water with  
207 much lower ORP (364 mV) but with the same ACC (i.e. < 0.01 mg/L) and comparable pH (i.e. 3.66). It  
208 was found that *E. coli* and *L. innocua* populations were reduced by  $0.97 \pm 0.07$  log CFU/mL and  $1.05 \pm$   
209  $0.03$  log CFU/mL, after one minute, respectively.

210

211

## DISCUSSION

212 Our study showed that the physicochemical properties of NEW with an ACC concentration of 480  
213 mg/mL were more stable at lower temperature (4 °C versus 25 °C) in closed (*cf.* opened) containers,  
214 and with lower SA:V ratios (0.9>1.7>8.7) over a storage period of 101 days (Fig. 1). The observed  
215 increase in stability of NEW at 4 °C is consistent with previous studies. For instance, Meireles *et al.*  
216 (17) reported that the ACC of NEW (with an initial ACC of 100 ppm) was more stable at 5°C than at  
217 25 and 30°C after 200 days. A similar observation was also made for Xin *et al.* (32) when NEW (with  
218 an initial ACC of 98 ppm, ORP of 875 mV and pH 6.5) was stored at 4 °C compared to 20 and 35 °C for  
219 over a period of three weeks.

220 The observed effects of temperature on NEW also agree well with those on acidic EO water.  
221 Previous studies showed that acidic EO water (pH <3) stored in glass bottles for up to 398 days was  
222 more stable at 4 °C than at  $\geq 20$  °C (6, 24). However, comparison of the observed shelf-stability of  
223 NEW with that previously observed for acidic EO water revealed that NEW might have a longer shelf-  
224 stability than acidic EO water (16, 24, 27), particularly at low storage temperatures and in sealed  
225 containers with low SA:V ratios. Of particular note was a long stability study (i.e., 398 days) that  
226 showed acidic EO water to be unstable at both 4 and 25 °C with ACC decreasing to <1 mg/L within  
227 the first 65 days (24). The longer shelf-stability of NEW than acidic EO water also agrees well with

228 the idea that the loss of ACC is due to volatilisation of the chlorine gas ( $\text{Cl}_2$ ) into the headspace. This  
229 reaction would most likely occur faster at higher temperatures and lower pH values; and potentially  
230 increase degradation of  $\text{HClO}$  under near neutral pH conditions at the higher temperatures *via* a  
231 chlorine dioxide intermediate with release of a proton (1). This might explain the observed changes  
232 in pH and ORP (Fig. 1b and 1c).

233 The effects of the SA:V ratio on the ACC of NEW were more noticeable at 25 °C than at 4 °C (Fig  
234 1a). The ACC decreased faster in bottles with an increase in the SA:V ratio ( $0.9 < 1.7 < 8.7$ ). To our  
235 knowledge, the effect of container shape on shelf stability has not been examined previously but our  
236 results suggest that the size of the headspace of containers may potentially effect shelf stability of  
237 NEW rather than SA:V ratio per say. In practical terms, shelf stability of NEW could be improved by  
238 storing it in containers that are filled to maximum capacity to minimise head space. Similarly, the  
239 physicochemical properties of NEW were more stable in closed bottles than in opened bottles (Fig.  
240 2). This is consistent with a previous study in which there were little changes in the ACC and ORP of  
241 NEW, acidic, or slightly acidic EO water in closed bottles and not in opened bottles when stored at  
242 20°C (3, 8, 33). . Taken together, the higher stability of EO water observed in bottles with lower SA:V  
243 ratio and in closed bottles observed might be because the evaporation rate of the dissolved  $\text{Cl}_2$   
244 increases with an increase in the SA:V ratio (1), thereby driving the decomposition of  $\text{HClO}$  in the EO  
245 water. Furthermore, the decomposition of  $\text{HClO}$  leads to hydrochloric acid and oxygen, resulting in a  
246 decrease in the pH (29).

247 In addition, aged EO water with different ACC and ORP levels but at the pH of  $3.4 \pm 0.2$  exhibited  
248 different antimicrobial effects on *E. coli* and *L. innocua* (Table 1). EO at an ACC of 63 mg/L and ORP of  
249 1,121 mV caused a  $>9$  log CFU/mL reduction of both test organisms within one minute at  $\sim 20$  °C. In  
250 contrast, EO water with low ACC (7 ppm) but similar ORP (1,094 mV) was able to reduce *E. coli* by  
251 approximately half ( $\sim 4.8$ -log reduction). EO water with a much lower ACC and ORP (ACC of  $< 0.01$   
252 mg/L and ORP of either 364 and 716 mV) produced the least antimicrobial effects, reducing both  
253 bacterial species by only  $\sim 1$  log CFU/mL within one minute. These results, taken together, indicate

254 that ACC plays a major role in the antimicrobial effects of EO water while pH and ORP contribute  
255 much less. Similarly, Waters & Hung (28), showed that EO water with pH of 2.8 and ACC of 0 mg/L  
256 had little effects on *E. coli* (<0.3-log reduction), whereas treatment with EO water with pH of 7.0 but  
257 higher ACC (34 mg/L) caused a 4.8-log reduction of *E. coli*. It also should be noted that the lack of  
258 efficacy of EO water with non-detectable ACC, but considerably high ORP (716 mV), was inconsistent  
259 with previously published data which ORP values of  $\geq 650$  mV were effective in eliminating bacteria  
260 within a few seconds although ACC was not reported (26, 34).

261 We determined the effects of various storage conditions on the stability of NEW in the context of  
262 commercial use (i.e., with high ACC) as a sanitiser. Our data revealed that the stability of the  
263 physicochemical properties of NEW (i.e., ACC, pH and ORP) was higher at low temperature (4 °C  
264 versus 25 °C) in a closed container and with a low SA:V ratio (0.9>1.7>8.7). Results from this study  
265 indicate that the antimicrobial effects of aged EO water is primarily dependent upon ACC rather than  
266 ORP and pH. The results of this study provide a useful insight for the food industry to accumulate  
267 and store NEW under the conditions that best preserve its stability as a sanitizer (i.e., maintaining  
268 ACC). However, further experiments to measure the stability of NEW when produced and stored on  
269 an industrial scale (e.g., 1,000 – 10,000 L closed polyethylene holding tanks of NEW at 500 mg/L ACC)  
270 for use as a sanitiser should be considered. A predictive model for NEW stability should also be  
271 developed to provide a tool for industry to better manage EO water.

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277 for valuable comments on an earlier draft of the manuscript.

278

279 **Table S1.** Summary of analysis of variance of Experiment 1 showing the effects of volume,  
 280 temperature and sampling day, and their interactions on ACC, ORP and pH of NEW stored for up to  
 281 101 days.

282 **pH**

283 Effect	Num DF	Den DF	F Value	P Value
284 Volume	2	6	50.63	0.0002
285 Days	17	210	16976.0	<.0001
286 Volume × Days	34	210	67.92	<.0001
287 Temp	1	210	300354	<.0001
288 Temp × Volume	2	210	55.20	<.0001
289 Temp × Days	17	210	10585.2	<.0001
290 Temp × Volume × Days	34	210	57.49	<.0001

291

292 **ORP**

293 Effect	DF	DF	F Value	Pr > F
294 Volume	2	6	4257.57	<.0001
295 Days	17	210	313.82	<.0001
296 Volume × Days	34	210	856.94	<.0001
297 Temp	1	210	194.15	<.0001
298 Temp × Volume	2	210	7690.14	<.0001
299 Temp × Days	17	210	307.41	<.0001
300 Temp × Volume × Days	34	210	866.43	<.0001

301

302 **ACC**

303 Effect	Num DF	Den DF	F Value	P Value
304 Volume	2	23.2	3.01	0.0686
305 Days	1	306	1490.38	<0.001
306 Days × Volume	2	306	7.39	0.0007
307 Temp	1	306	61.08	<0.001
308 Temp × Volume	2	306	2.92	0.0556

309	Days × Temp	1	306	460.08	<0.001
310	Days × Temp × Volume	2	306	1.71	0.1827

311  
312

313 **Table S2.** Summary of analysis of variance of Experiment 2 showing the effects of treatment (open vs  
314 closed), sampling day, and their interactions on ACC, ORP and pH of NEW stored for up to 101 days.

315 **pH**

316	Effect	Num DF	Den DF	F Value	P Value
317	Treatment	1	30	271.02	<.0001
318	Days	7	30	2161.16	<.0001
319	Treatment × Days	7	30	80.56	<.0001

320

321 **ORP**

322	Effect	Num DF	Den DF	F Value	P Value
323	Treatment	1	30	7.14	0.0105
324	Days	7	30	695.79	<.0001
325	Treatment × Days	7	30	40.57	<.0001

326

327 **ACC**

328	Effect	DF	DF	F Value	Pr > F
329	Treatment	1	30	0.00	0.9961
330	Days	7	30	314.29	<.0001
331	Treatment × Days	7	30	5.39	0.0249

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334 **Figure S1.** Representative bottles showing surface-area to volume ratios (SA:V 8.7, 1.7 and 0.9) used  
335 to store the neutral electrolysed water.

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- 428

429 Fig. 1: Effect of temperature, 4 °C (black) and 25 °C (grey) on A) concentration of available chlorine  
430 (ACC), B) pH and C) ORP of NEW with SA:V ratios of 8.7 (square), 1.7 (circle), and 0.9 (triangle) stored  
431 for up to 101 days. Error bars equal to 1 S.D.

432

433 Fig. 2: Effect of 'open' (black) or 'closed' (grey) containers on the rate of change of the A)  
434 concentration of available chlorine B) pH and C) the ORP of NEW stored at 25 °C overtime. Error bars  
435 equal to 1 S.D.

436 TABLE 1. Antimicrobial effects of stored EO water on *Escherichia coli* and *Listeria innocua*. Different  
 437 letters (a, b, c) denote a significant treatment ( $\alpha = 0.05$ ) effect within each exposure time. One S.D is  
 438 shown in parentheses. n=3 for each EO water treatment within each exposure time.  
 439

				Mean Log reduction <sup>‡</sup> (log CFU/mL)		
				Exposure time (minutes)		
Bacteria	ACC (mg/L)	pH	ORP (mV)	1	5	10
<i>E. coli</i>	<0.01*	3.66	364	0.97 (0.07) a	1.03 (0.04) a	0.96 (0.05) a
	<0.01*	3.24	716	0.98 (0.08) a	0.93 (0.08) a	0.94 (0.07) a
	7	3.35	1,094	4.76 (0.09) b	4.81 (0.07) b	4.72 (0.13) b
	63	3.26	1,121	>9 <sup>+</sup> (0.00) c	>9 <sup>+</sup> (0.00) c	>9 <sup>+</sup> (0.00) c
<i>L. innocua</i>	<0.01*	3.66	364	1.05 (0.03) a	1.01 (0.02) a	1.05 (0.02) a
	<0.01*	3.24	716	1.02 (0.02) a	0.99 (0.04) a	0.99 (0.02) a
	63	3.26	1,121	> 9 (0.00) b	>9 <sup>+</sup> (0.00) b	>9 (0.00) b

440 ‡) Mean Log reduction = initial counts – final count at each sampling time point (data not shown)

441 \*) Below detection limit of Compact ClO<sub>2</sub>+ meter (0.01 mg/L)

442 +) Below detection limit of enumeration method < 1 log CFU/mL

443 ACC, available chlorine concentration.

444 ORP, oxidation reduction potential.

445